Abstract

Background: Breast cancer is the main cause of cancer in women and the second cause of malignancy deaths. Ki-67 is one of the molecular markers used to evaluate cancer prognosis along with other factors such as age, tumor size, lymph node involvement, estrogen receptor (ER), progesterone receptor (PR), P53, human epidermal growth factor receptor-2 (HER-2), histological and nuclear grades. This study was aimed to evaluate the correlation of KI-67 expression with some biomarkers and clinico-pathological characteristics in breast cancer patients. Materials and Methods: A total of 513 cases (all female) aged 40-80 years, were randomly selected from patients who were admitted in two centers affiliated with Tehran University of Medical Sciences (Buoo-alli and Kasra hospitals) over a 7-year period (2010-2015). Assessment of tumors for HER-2, P53, ER PR, pathological type and histologic grade was performed. Ki-67 labelling index (Ki-67LI) was defined as the percentage of MIB1-positive cells among a total number of 1,000 malignant cells at high-power magnification (×400). Results: Our study showed that age, ER and PR status were negatively correlated with Ki-67LI (P<0.05). Moreover, number of lymph nodes involved, HER-2, P53 and nuclear grades had a positive correlation with Ki-67LI (P<0.05), whereas, tumor size and histological grade showed no significant correlation with Ki-67LI (P=0.195 and P=0.721, respectively). Conclusion: Results of our study and other studies confirm that the expression of Ki-67 is significantly associated with ER, PR, HER-2 and P53 status. On the other hand, Ki-67 relationship with clinical characteristics such as age, tumor size and lymph node metastasis is not completely established and needs further research. [GMJ.2016;5(2):90-97]

Keywords: Breast Cancer; Ki-67; Estrogen Receptor; Progesterone Receptor; P53; Human Epidermal Growth Factor Receptor-2
Introduction

Breast cancer is a major public health issue and chief cause of cancer and cancer-related mortality in women in Iran and many other parts of the world [1]. It is also the fifth cause of death in females in Iran [2]. Incidence of breast cancer varies around the world, it is more common in wealthy countries due to higher existence of risk factors such as higher age at first pregnancy, high calorie intake, sedentary lifestyle and the use of hormonal contraceptives [3,4]. Breast cancer prognosis is worse in less wealthy countries and in patients with lower income or educational level [4]. Although, it is of lowest incident in Asian countries; this rate is rising [5]. Mean age of breast cancer in Iran is 49 years old, whereas its mean age in western countries is 10 years older [6,7].

Age, tumor size, lymph node involvement, histological and nuclear grades of tumor are some of the prognostic factors for breast cancer [8]. In the past years, molecular research has allowed using different molecular markers including estrogen and progesterone receptors (ER and PR), human epidermal growth factor 2 (HER-2), P53 and recently added Ki-67 to predict the disease outcome. Expression of these markers impact the course of disease and treatment plan, since tumors with positive ER and PR status correspond better to hormone therapy and positive HER-2 tumors can be treated with Trastuzumab [9-10].

Ki-67 antigen, originally found by Gredes et al. [11], is involved in the initial steps of polymerase I dependent ribosomal RNA production [12]. Cells express this antigen during all cell-cycle phases except G0 (the resting phase) [13]. Ki-67 levels are highest in early mitosis; in later mitotic phases (anaphase and telophase), its levels sharply decrease [14]. Ki-67 is associated with mitotic activity of the tissue, thus it correlates with mitotic index and because Ki-67 can be detected easier than mitotic index, it is thought to be a rather superior prognostic marker [15]. Trihia et al. used Ki-67 as an alternative for mitotic index in Nottingham grade and produced Nottingham/Ki-67 grade, and when it was evaluated for prognostic significance, it was similar to Bloom–Richardson grade and Nottingham grade [16]. Weidnet et al. measured Ki-67 using MIB-1 antibody on paraffin embedded breast carcinoma samples and concluded that Ki-67 strongly correlates with both mitotic figure count and tumor grade [17]. Ki-67 is often measured on paraffin embedded sections using the MIB-1 antibody-based on percentage of tumor cells stained by the antibody. Normal breast tissue can express Ki-67 less than 3% [18].

Ki-67 is studied mainly for its prognostic role in breast cancer; however, research continues to find other roles such as predicting the outcome of certain treatments, or identifying patients eligible for a certain adjuvant-therapy regime. A study on 3652 breast cancer patients in Japan revealed that higher Ki-67 profile significantly correlated with higher grade of malignancy, lower disease-free survival and overall survival, poorer prognosis and early recurrence [19]. A meta-analysis of 46 studies confirmed that Ki-67 was positivity associated with higher relapse and worse survival in breast cancer patients [20]. From 105 triple negative (ER, PR and HER2 negative) breast cancer patients under treatment of Doxorubicin and Docetaxel, those with higher Ki-67 (≥10%) responded better to treatment, but had lower relapse-free survival and overall survival [21]. Another study demonstrated that Ki-67 improved the prediction of treatment response and prognosis in 552 breast cancer patients receiving neoadjuvant treatment, as mean Ki-67 values in patients with a pathological complete response were higher than patients without a complete response [22]. Considering the importance of Ki-67 in breast cancer, this study was aimed to evaluate the correlation of Ki-67 expression with some biomarkers and clinicopathological characteristics in breast cancer patients.

Materials and Methods

1. Patients and Tumor Samples

This was a cross-sectional study done on female patients older than 18 years referring to Buali or Kasra hospitals in Tehran, Iran between 2010 and 2015 for breast biopsy or surgery, and were diagnosed with breast can-
cer in their pathology report. Total number of patients was 526 from whom 9 were males, 3 had bilateral tumor and one patient had a history of left mastectomy despite present tumor in right breast; these were excluded and the remainder of 513 patients entered the study. No calculation of sample size was done, and all qualified patients entered the study.

2. Clinicopathological Features
Data regarding age, tumor size, lymph node involvement, nuclear grade and histological grade were extracted from patients’ records.

3. Immunohistochemistry (IHC) Assessment
All samples were evaluated by (IHC) staining under the direct supervision of at least two pathology academics.

3.1. Assessment of Ki-67
Formalin-fixed paraffin tissue sections (FFSs, 4 μm) mounted on Superfrost slides (Superfrost) were IHC stained, by using the standard streptavidin-biotin complex method, as previously described [23]. Microwave-assisted heat-induced retrieval method for antigen epitopes was performed in citrate buffer, at pH 6.0 for 20 minutes. Endogenous peroxidase activity was blocked by incubation in a 0.3% hydrogen peroxide in methanol buffer for 10 minutes. Nonspecific binding of primary antibody was blocked by using normal swine serum (NSS, in Tris-buffered saline (TBS) (1:5), 100 μl/slide) for 10 minutes of incubation. Primary mouse monoclonal anti-Ki-67 antibody (MIB1 clone, product M7240; Dako, Glostrup, Denmark), diluted 1:100 (optimum working dilution) in NSS/TBS, was applied to each slide and incubated for 60 minutes at room temperature. Slides were then rinsed in TBS before staining with a streptavidin-biotin three-stage technique, with Dako Strept ABC complex/HRP Duet kit (Dako, K492) according to manufacturer’s guidelines. For reaction visualization, 3-3′-diaminobenzidine tetrahydrochloride (Dako liquid DAB Plus, K3468) was used as chromogen. The sections were counterstained with Mayer hematoxylin (Dako, AR106). Human tonsil sections were used as positive control, whereas negative control was performed by replacing the primary antibody by TBS.

3.2. Ki67 Scoring
Immunostaining was quantitatively evaluated by using light microscopy, in which the entire section was scanned at low-power magnification (×100) to determine areas with the largest number of positive nuclei (hot spot) within the invasive component [23]. These were usually found at the periphery of tumors and were easier to identify than the mitotic figure hot spots. Ki-67 labeling index (Ki-67LI) was expressed as the percentage of MIB1-positive cells among a total number of 1,000 malignant cells at high-power magnification (×400).

3.3. Determining HER-2 Status
HER-2 status was determined by means of IHC using Dako Hercep Test (Dako, Copenhagen, Denmark) and scored with Dako scoring system [9]. Only patients who had weak-to-moderate staining of the entire tumor-cell membrane for HER-2 (referred to as a score of 2+) or more than moderate staining (referred to as a score of 3+) in more than 10 percent of tumor cells on IHC analysis were eligible for the study.

3.4. Determining ER and PR Status
ER and PR status were determined with a modified avidin-biotin (ABC) immunoperoxidase method according to standard protocols (Vector Laboratories, Burlingame, CA). 3,3′-diaminobenzidine was used as chromogen. The immunostaining results for ER and PR were assessed semi-quantitatively and reported as positive if more than 5% of cells were immunostained in a tumor.

3.5. Determining P53 Overexpression
P53 overexpression was defined as more than 50% of the cells with strong nuclear staining as previously described [9].

4. Statistical Analysis
Due to normal distribution of selected variables evaluated by Kolmogorov-Smirnov test, we used simple linear regression model and Pearson correlation coefficient to evaluate the association between tumor size, number of involved lymph nodes, patients’ age, histological grade, nuclear grade, ER, PR, P53 and HER-2 with Ki-67LI. Data was analyzed using SPSS version 20; P-values smaller than 0.05 were defined as significant.
Results

Mean age of patients enrolled in this study was 52.8±11.71 years. Moreover, maximum and minimum tumor sizes were 17cm and 0.3cm, respectively, and mean tumor size was 2.9±1.77cm. We distributed patients according to their age into 6 groups. Most patients were in 40-50 years range, mean expression of Ki-67LI was measured for each group; patients aged 30-40 years had the highest mean Ki-67LI (34.17%) and patients between 60-70 years old had the lowest mean Ki-67LI (27.22%). Most of the cases had a tumor size between 1cm and 3cm (64.33%), mean Ki-67LI was highest in tumors between 5cm and 7cm (35.09%) and lowest in lower than 1cm tumors (27.93%), considering the reported tumor sizes. Lymph node involvement was not observed in 36.65% of patients, moreover, mean Ki-67LI was maximal in patients which had 7 to 9 of their lymph nodes involved. Histological grade was reported using Scarff-Bloom-Richardson grading system, better differentiated tumors have lower histological grade. More than half of subjects (54.58%) had a histological grade of 2 and mean Ki-67LI increased along with histological grade, with Ki-67LI of 36.52% in grade 3. Sixty-three point ninety-four percent of patients had a nuclear grade of 0 and patients with nuclear grade of 1 had the lowest mean Ki-67LI (23.18%); highest mean Ki-67LI (40.87%) was observed in nuclear grade of 3. ER and PR positive patients had lower Ki-67LI (28.37% and 29.56%, respectively), whereas P53 and HER-2 positive patients had higher Ki-67LI (36.27% and 35.45%, respectively) (Table-1).

Considering normal distribution of tumor size, number of involved lymph nodes, age, histological grade, nuclear grade, ER, PR, P53 and HER-2, we used simple linear regression model and Pearson correlation coefficient to evaluate their associations with Ki-67LI. Table-2 shows the results of linear regression model; from selected variables tumor size and histological grade showed no significant correlation with Ki-67LI (P=0.195 and P=0.721, respectively). Age, ER and PR had inverse associations with Ki-67LI expression. (P< 0.05)
On the contrary, Sahin et al. found a strong correlation between Ki-67 staining percentage and age in addition to correlation with nuclear grade, mitotic rate and 5-year disease free and overall survivals [25]. Moreover, a study of 203 female breast cancer patients revealed that Ki-67 statistically correlated with tumor size and nuclear estrogen receptor content, and premenopausal women had greater Ki-67 values (median value, 14.1%) than postmenopausal ones (median value, 9.8%). Similar to Veronese et al. and Crispino studies, no correlation with lymph node involvement was found [24, 26]. Interestingly, a study in Iran demonstrated significant association between axillary lymph node involvement and level of Ki-67 [12].

P53 is another tumor marker and prognostic factor used in breast cancer; several studies tried to find the associations between P53 and other markers including Ki-67 as well as comparing them to find the superior prognostic factor. A study on 71 primary breast carcinoma specimens revealed that p53 protein was associated with high levels of Ki-67 and lymph-node status [27]. Ki-67 was correlated with high mitotic count, histologic grade, negative progesterone receptor status and P53 expression in another study done on 97 breast carcinomas [27]. Li et al. studied 151 cases of breast cancer and found that Ki-67 did not relate to age, tumor size and lymph node status, whereas it correlated with tumor stage and P53 expression [28]. Two studies in Iran compared the prognostic value of P53 with Ki-67 in breast cancer and achieved contradictory results [29, 30]. The study in western Iran showed that the hazard ratios for P53 and Ki-67 were 1.37 and 0.52, respectively. Thus, P53 is more important than Ki-67 on survival rate [29]. Another study in eastern Iran demonstrated that Ki-67 had significant relationship with the survival rate, but over-expression of P53 did not show such a significance. Hence, they concluded that Ki-67 marker is more important than P53 protein in breast cancer prognosis [30].
Table 2. Correlation Between Selected Variables and Ki-67 Using Simple Linear Regression

<table>
<thead>
<tr>
<th>Variable</th>
<th>B¹</th>
<th>95% Confidence interval</th>
<th>Standard deviation</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Lower</td>
<td>Upper</td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td>-0.150</td>
<td>1.74</td>
<td>3.66</td>
<td>0.078</td>
</tr>
<tr>
<td>Tumor size</td>
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<td>1.34</td>
<td>5.26</td>
<td>0.629</td>
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<tr>
<td>Lymph nodes involvement</td>
<td>0.231</td>
<td>1.64</td>
<td>3.14</td>
<td>0.087</td>
</tr>
<tr>
<td>ER</td>
<td>-9.032</td>
<td>-5.34</td>
<td>2.93</td>
<td>2.291</td>
</tr>
<tr>
<td>PR</td>
<td>-3.951</td>
<td>-2.92</td>
<td>2.12</td>
<td>2.082</td>
</tr>
<tr>
<td>HER-2</td>
<td>3.282</td>
<td>1.88</td>
<td>8.92</td>
<td>1.193</td>
</tr>
<tr>
<td>P53</td>
<td>3.878</td>
<td>1.43</td>
<td>2.49</td>
<td>1.244</td>
</tr>
<tr>
<td>Histological grade</td>
<td>0.855</td>
<td>-4.16</td>
<td>6.16</td>
<td>2.293</td>
</tr>
<tr>
<td>Nuclear Grade</td>
<td>6.117</td>
<td>2.84</td>
<td>9.16</td>
<td>2.182</td>
</tr>
</tbody>
</table>

¹ B: regression coefficient for dependent variable
ER: Estrogen receptor, PR: Progesterone receptor, HER-2: Human epidermal growth factor receptor-2

Conclusion

Despite similar results regarding Ki-67 correlation with tumor grade, P53, ER and PR there is still no uniform opinion in terms of Ki-67 association with other clinical characteristics such as age, tumor size or lymph node involvement. These results vary due to different types of studied breast cancer samples or sample size population characteristics, and a consonant conclusion may never be found in general population. What is most certain about Ki-67 is its role in breast cancer prognosis, and future studies should focus more on finding other functional values for Ki-67 such as treatment prediction and cancer drug therapy.

Conflict of Interest

The authors declare that they have no conflicts of interest.

References


23. Aleskandarany MA, Green AR,


